DRUG DISCOVERY

16(37), 2022

Renal impact of desloratadine/dihydroartemisinin /piperaquine on healthy and parasitized mice

Georgewill UO1, Ebong NO1, Adikwu E2*

To Cite:

Georgewill UO, Ebong NO, Adikwu E. Renal impact of desloratadine/dihydroartemisinin/piperaquine on healthy and parasitized mice. *Drug Discovery*, 2022, 16(37), 45-52

Author Affiliation:

Department of Pharmacology, Faculty of Basic Clinical Sciences, University of Port Harcourt, Rivers State, Nigeria.

Department of Pharmacology/Toxicology, Faculty of Pharmacy, Niger Delta University, Bayelsa State, Nigeria.

*Corresponding author:

Department of Pharmacology/Toxicology, Faculty of Pharmacy, Niger Delta University, Bayelsa State, Nigeria Email: adikwuelias@gmail.com

Peer-Review History

Received: 05 February 2022 Reviewed & Revised: 12/February/2022 to 06/April/2022 Accepted: 08 April 2022 Published: 11 April 2022

Peer-review

External peer-review was done through double-blind method.



© The Author(s) 2022. Open Access. This article is licensed under a Creative Commons Attribution License 4.0 (CC BY 4.0)., which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/.

ABSTRACT

Desloratadine/dihydroartemisinin/piperaquine (DL/D/P) showed promising therapeutic activity on Plasmodium berghei. This study evaluated its renal impact on healthy and Plasmodium berghei-infected mice. Fifty-four adult Swiss albino mice used were randomized into 9 groups. Thirty mice (n=6/group) were inoculated with Plasmodium berghei (1 X 107) and treated with normal saline (0.2ml) (Parasitized control), DL, D/P and DL/D/P daily for 4 days, respectively. The non-parasitized control was treated with normal saline (0.2ml) daily for 4 days. In the sub-acute toxicity study, twenty four healthy mice (n=6/group) were treated with normal saline (0.2ml) (Control), DL, D/P and DL/D/P daily for 28 days, respectively. After treatment, the mice were weighed and anesthetized. Blood samples were collected and evaluate for renal biochemical markers. Kidney samples were weighed and analysed for markers of oxidative stress and histology. DL, D/P and DL/D/P did not produce significant (p>0.05) effects on renal function markers in parasitized mice when compared to control. DL, D/P and DL/D/P significantly decreased body weight and significantly increased kidney weight in healthy mice at p<0.05, p<0.05 and p<0.01, respectively when compared to control. Serum creatinine, urea, uric acid and kidney malondialdehyde levels were increased significantly in DL (p<0.05), D/P (p<0.05) and DL/D/P (p<0.01) treated healthy mice when compared to control. Significantly decreased kidney glutathione peroxidase, superoxide dismutase, glutathione, and catalase levels were observed in healthy mice treated with DL (p<0.05), D/P (p<0.01) and DL/D/P (p<0.001) when compared to control. DL/D/P produced tubular necrosis, vacuolated glomerular mesangial cells and increased Bowman's space in healthy mice. The prolonged use of DL/D/P may cause renal dysfunction.

Keywords: Dihydroartemisinin/piperaquine, Desloratadine, *Plasmodium*, Mice, Renal, Toxicity

1. INTRODUCTION

The kidney is essential for the disposal of waste products, regulation of extracellular fluid volume, electrolyte concentrations, and serum osmolality



(Bhatt and Jialal, 2020). Allopathic medicine has subjected patients to taking a variety of drugs for diagnostic and therapeutic reasons (Pazhayattil and Shirali, 2014). The kidneys provide the ultimate solution for the excretion and elimination of many drugs and metabolites, which often subject them to elevated levels of potentially toxic substances. As a result, kidney tubular cells and papillae are exposed to direct toxic damage causing nephrotoxicity (Mahmoudi et al., 2021). Drugs can cause nephrotoxic complications such as interstitial nephritis, nephrotic syndrome, impaired intraglomerular hemodynamics, tubulointerstitial disease, and renal scarring resulting in acute or chronic kidney injury (Sari, 2019). Nephrotoxic complications associated with drugs can by heralded by alterations in indices such glomerular filtration rate, blood urea nitrogen, serum creatinine, or urine output; however, some indices may remain unperturbed (Lillie and Cummings, 2018).

Nephrotoxicity associated with antimalarial drugs may be rare, but there are growing concerns based on reported observations (Wiwanitkit, 2015). Chloroquine may alter kidney structure and also impair kidney function causing inappropriate retention of sodium and chloride in renal tubules and alterations in renally active hormones (Musabayane et al., 2000). Quinine can precipitate acute kidney injury through immune-mediated reactions, especially thrombotic microangiopathy (Al-nouri et al., 2015; Liles et al., 2015). Artesunate may cause nephrotoxicity marked by diminished glomerular filtration capacity, increased kidney blood flow and urinary excretion of electrolytes (Campos et al., 2001) which may be reversible as reported in animal studies (Li et al., 2007).

Dihydroartermisinin-piperaquine (D/P) is recommended by the World Health Organization (WHO) for the treatment of *Plasmodium falciparum* (*P.falciparum*) associated malaria (WHO, 2015). The complementary mechanisms of action of the combined drugs increase the effectiveness of treatment and prevent or delay the emergence of resistance (Reuter at al., 2015). D/P has demonstrated 100% clinical efficacy for the treatment of uncomplicated *P. falciparum* malaria with a day-3 parasitemia-positive rate of 6.2% (Sun et al., 2011). Nevertheless, the development of *Plasmodium* parasite resistance was observed in malaria endemic regions (Georgewill et al., 2021) and the possible occurrence of nephrotoxicity (Alabi et al., 2018). Desloratadine (DL) is a second-generation non-sedating antihistamine with long-acting activity used for the treatment of seasonal allergic rhinitis and idiopathic urticarial (Kazmi et al., 2015). In addition to its anti-histamine activity, it has shown promising antimalarial activity (Aneesa, 2011). DL has been shown to increase the antiplasmodial activity of D/P, which suggests possible clinical use as an antimalarial drug (Georgewill et al., 2021). However, there is lack of scientific safety data on DL and D/P combination. The current study deemed it imperative to assess the safety of DL/D/P by tacking into cognisance effect on the kidneys of healthy and *P. berghei*-infected mice.

2. MATERIALS AND METHODS

2.1. Drugs, Animals, and Malaria Parasite

Fifty-four adult Swiss mice (22-25g) were procured from the animal unit of the Department of Pharmacology, Faculty of Basic Cinical Sciences, University of Port Harcourt, Rivers State, Nigeria. The mice were grouped (n=6/group) and acclimated for 2 weeks before the study commenced. The mice were kept under natural conditions and had access to food and water freely. Desloratadine (DL) (Merck & Co), and Dihydroartemisinin/piperaquine (D/P) (Bliss GVS Pharma Ltd India) were used. The doses of (D/P) (1.71/13.7 mg/kg), and DL (5 mg/kg) used were derived from previous antiplasmodial studies (Georgewill et al., 2021).

2.2. Parasite inoculation of mice and treatment

Mice infected with chloroquine-sensitive P. berghei (NK65) supplied by the Nigerian Institute of Medical Research (NIMR), Yaba, Lagos, Nigeria, were used as the donor. Thirty mice were randomised into 5 groups of n=6/group. Groups II-V were inoculated intraperitoneally (i.p) with P. berghei containing 1×10^7 parasitized erythrocytes. After 3 days, treatment commenced orally as follows: Group 1(Normal control) and group II (parasitized control): normal saline (0.2 ml), groups III-V: (D/P) (1.71/13.7 mg/kg), DL (5 mg/kg) and DL/D/P daily for 4 days, respectively.

2.3. Treatment of healthy mice

Twenty-four adult Swiss albino mice were grouped into 4 of n=6/group. Group I (Control) daily received normal saline (0.2mL) orally for 28 days. Groups II-IV orally received (D/P) (1.71/13.7 mg/kg), DL (5 mg/kg) and DL/D/P daily for 28 days, respectively.

2.4. Animal sacrifice

After treatment, the mice were fasted overnight, weighed and anesthetized (diethylether), and blood samples were obtained by cardiac puncture. Blood samples were centrifuged (1200 rpm for 20 minutes) and sera separated and evaluated for renal biochemical markers. The mice were sacrificed, kidneys were harvested, rinsed in saline and homogenized in 0.1 M Tris-HCl

solution buffered (pH 7.4). The homogenates were centrifuged (2000 rmp for 20 minutes) and the supernatants decanted and evaluated for oxidative stress markers.

2.5. Serum biochemical markers assessments

Sera were evaluated for creatinine, urea, uric acid, and electrolytes (sodium, potassium, chloride, and bicarbonate ions) using an auto analyser.

2.6. Oxidative stress marker assay

Kidney glutathione (GSH) was assessed as described by Sedlak and Lindsay (1968). Catalase (CAT) was assessed as explained by Aebi, (1984). Glutathione peroxidase (GPx) was evaluated according to the protocol reported by Rotruck *et al.* (1973). Superoxide dismutase (SOD) was determined as described by Sun and Zigman (1978). Malondialdehyde (MDA) was measured using the protocol explained by Buege and Aust (1978).

2.6. Histology of the kidney

Harvested kidney tissues were cut and immersed in Bouin's solution for 24hr. The tissues were dehydrated in alcohol-graded series, processed and fixed in paraffin wax. Sections (3 μ m) were cut and stained (Haematoxylin and Eosin) on slides. The slides were examined under light microscope and appropriate sections photographed using a digital camera.

2.7. Statistical analysis

Data are presented as mean ± standard error of mean (SEM). Differences between groups were assessed using one-way analysis of variance (ANOVA) followed by Tukey's multiple range test (Graph Pad Prism 5 Software, San Diego, CA USA). *P* values less than 0.05, 0.01 and 0.001 were considered statistically significant.

3. RESULTS

3.1. Effects of desloratadine/dihydroartemisinin/piperaquine on body, kidney weights and serum biochemical markers of parasitized mice

Parasitized mice treated with DL, D/P and DL/D/P for 4 days showed no evident (P > 0.05) changes in body and kidney weights when compared to control (Table 1). Treatment of parasitized mice with DL, D/P and DL/D/P for 4 days had no significant (P > 0.05) effects on serum urea, uric acid, creatinine and serum electrolytes when compared to control (Tables 2 and 3).

3.2. Effects of desloratadine/dihydroartemisinin/piperaquine on body, kidney weights and serum biomarker of healthy mice

Healthy mice treated with DL, D/P and DL/D/P for 28 days showed significantly reduced body weight at P < 0.05, P < 0.05 and P < 0.01, respectively when compared to control (Table 1). Treatment of health mice with DL, D/P and DL/D/P significant increased kidney weight at P < 0.05, P < 0.05 and P < 0.01, respectively when compared to control (Table 1). Serum creatinine, urea, and uric acid levels were elevated significantly in healthy mice treated with DL (P < 0.05), D/P (P < 0.05) and DL/D/P (P < 0.01) when compared to control (Table 2). The effects of DL, D/P and DL/D/P on serum electrolytes levels in healthy mice were not significantly (P > 0.05) different from the control (Table 3).

Table 1: Effects of deslorated ine/dihydroartemisinin/piperaquine on body and kidney weights of healthy and parasitized mice

Treatment	Final body weight (g)		Absolute kidn	ey weight (g)	Relative kidney weight (%)	
	Healthy mice	Parasitized mice	Healthy mice	Parasitized mice	Healthy mice	Parasitized mice
Control	30.10±2.18	29.60±3.01	0.58 ± 0.04	0.46±0.05	1.92±0.03	1.55±0.03
DL	24.20±2.72*	29.20±2.14	0.74±0.03*	0.43 ± 0.03	3.05±0.05*	1.47±0.07
D/P	23.70±2.23*	27.40±2.28	0.80±0.05*	0.44 ± 0.07	3.37±0.03*	1.61±0.05
DL/ D/ P	$20.20 \pm 3.47^{\pi}$	26.80±2.15	$0.99\pm0.02^{\pi}$	0.42±0.04	$4.90\pm0.04^{\pi}$	1.57±0.04

Data as mean \pm SEM, SEM: Standard error of mean. n=6, DL: Desloratadine, D/P: Dihydroartemisinin/piperaquine, * p<0.05, π p<0.01 Differ significantly when compared to control (Healthy mice) (ANOVA).

Table 2: Effect of desloratadine/dihydroartemisinin/piperaquine on serum renal function markers of healthy and parasitized mice

Treatment	Healthy mice		Parasitized mice			
	Creatinine (mg/dL)	Urea(mg/dL)	Uric acid (mg/dL)	Creatinine (mg/dL)	Urea (mg/dL)	Uric acid (mg/dL)
Control	0.72±0.05	9.35±0.07	3.84±0.20	0.78±0.04	9.77±0.03	1.58±0.34
DL	1.38±0.03*	12.50±1.15*	2.96±0.11*	0.73±0.06	9.86±0.06	1.64±0.28
D/P	1.51±0.07*	14.20±0.22*	3.57±0.09*	0.76±0.09	9.97±0.09	1.57±0.51
DL/D/P	$2.58\pm0.02^{\pi}$	$21.50\pm1.75^{\pi}$	$5.12\pm0.67^{\pi}$	0.78±0.05	9.99±0.05	1.53±0.69

Data as mean \pm SEM, SEM: Standard error of mean. n=6, DL: Desloratadine, D/P: Dihydroartemisinin/piperaquine, * p<0.05, π p<0.01 Differ significantly when compared to control (Healthy mice) (ANOVA).

Table 3: Effect of desloratadine/dihydroartemisinin/piperaquine serum electrolytes of healthy and parasitized mice

	Healthy mice					Parasitized	mice	_
Treatment	K	Na	C1 l	HCO ₃	K	Na	Cl	HCO ₃
	(mmol/L) (mmol/L)		(mmol/L) (mmol/L)		(mmol/L)	(mmol/L)	(mmol/L)	(mmol/L)
Control	5.84±0.72	121.76±13.2	135.25±14.1	14.25±1.86	5.57±0.39	107.96±12.9	123.87±14.0	15.27±1.80
DL	5.81±0.66	118.53±11.5	132.93±12.2	14.11±1.57	5.60±0.24	109.85±12.4	125.95±13.3	15.58±1.29
D/P	5.78±0.46	117.15±14.8	130.87±12.0	13.88±1.78	5.63±0.16	112.68±13.6	128.06±12.1	15.73±1.85
DL/D/P	5.76±041	115.43±11.6	127.74±13.4	13.65±1.39	5.67±0.21	114.66±12.1	130.80±10.7	15.86±1.55

Data as mean ± SEM, SEM: Standard error of mean. n=6, DL: Desloratadine, D/P: Dihydroartemisinin/piperaquine (ANOVA)

Table 4: Effect of desloratadine/dihydroartemisinin/piperaquine on kidney oxidative stress markers of parasitized mice

Treatment	MDA nmole/mg protein	GSH µmole/mg protein	CAT U/mg protein	SOD U/mg protein	GPx U/mg protein
Control	0.22±0.05	12.00±1.00	23.47±1.44	15.00±2.00	16.71±1.00
PC	0.21±0.07	11.98±1.34	23.40±2.21	14.98±1.63	16.69±1.00
DL	0.23±0.01	11.95±1.21	23.31±2.32	14.96±1.52	16.65±1.26
D/P	0.25±0.04	12.21±1.31	23.29±3.20	15.00±1.27	16.70±1.34
DL/ D/ P	0.27±0.06	11.97±0.67	23.26±3.41	14.89±1.33	16.57±1.33 ^π

Data as mean ± SEM, n=6, SEM: Standard error of mean, DL: Desloratadine, D/P: Dihydroartemisinin/piperaquine, MDA: Malondialdehyde, GSH: Glutathione, CAT: Catalase, SOD: Superoxide dismutase, GPx: Glutathione peroxidase.

Table 5: Effect of desloratadine/dihydroartemisinin/piperaquine on kidney oxidative stress markers of healthy mice

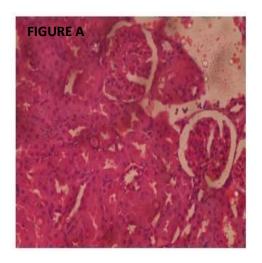
Treatment	MDA nmole/mg protein	GSH µmole/mg protein	CAT U/mg protein	SOD U/mg protein	GPx U/mg protein
Control	0.29±0.03	10.05±0.81	27.4±1.89	12.34±3.86	17.65±0.21
DL	0.49±0.06*	7.00±0.98*	21.3±2.15*	9.65±3.41*	13.20±0.27*
D/P	0.72±0.09**	5.12±0.53**	17.40±1.26**	7.73±1.57**	10.70±0.61**
DL/ D/ P	$1.47\pm0.02^{\pi}$	$4.23\pm0.39^{\pi}$	$12.90\pm0.83^{\pi}$	4.63±3.01 ^π	$7.52\pm0.25^{\pi}$

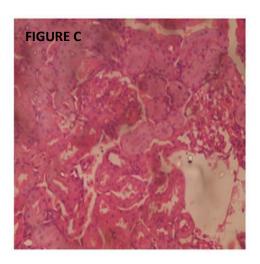
Data as mean ± SEM, n=6, SEM: Standard error of mean, DL: Desloratadine, D/P: Dihydroartemisinin/piperaquine, MDA: Malondialdehyde, GSH: Glutathione, CAT: Catalase, SOD: Superoxide dismutase, GPx: Glutathione peroxidase, * p<0.05, **p<0.01, π p<0.001 Differ significantly when compared to control.

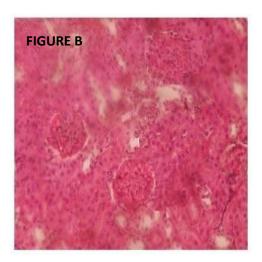
3.3. Effects of desloratadine/dihydroartemisinin/piperaquine on kidney oxidative stress markers of healthy and parasitized mice Kidney antioxidants (SOD, GPx, GSH and CAT) were unchanged (P > 0.05) in parasitized mice treated with DL, D/P and DL/D/P for 4 days when compared to control (Table 4). But kidney antioxidants were significantly reduced in healthy mice treated with DL (P < 0.05), D/P (P < 0.01) and DL/D/P (P < 0.001) for 28 days when compared to control (Table 5). MDA levels were unchanged (P > 0.05) in parasitized mice treated with DL, D/P and DL/D/P for 4 days (Table 4). In contrast, MDA levels were elevated in healthy mice treated with DL (P < 0.05), D/P (P < 0.01) and DL/D/P (P < 0.001) for 28 days when compared to control (Table 5).

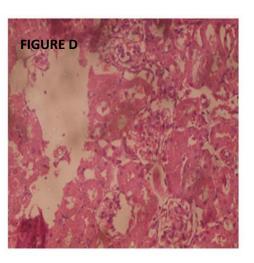
3.4. Effect of desloratadine/dihydroartemisinin/piperaquine on kidney histology of healthy mice

The kidney of the control mice showed normal glomerulus and renal tubules (Fig A) while the kidney of DL (Fig B) and D/P (Fig C) treated health mice showed normal renal tubules and vacuolated glomerular mesangial cells. Kidney of DL/D/P treated healthy mice showed tubular necrosis, vacuolated glomerular mesangial cells and widened Bowman's space (Fig D).









The kidney of control mice showed normal glomerulus and renal tubules (Figure A). Kidney of DL (Figure B) and D/P (Figure C) treated health mice showed normal renal tubules and vacuolated glomerular mesangial cells. Kidney of DL/D/P treated healthy mice showed tubular necrosis, vacuolated glomerular mesangial cells and widened Bowman's space (Fig D). H and EX 400

4. DISCUSSION

Studies showed promising antimalarial activity of DL/D/P (Georgewill et al., 2021), but without scientific literature on its toxicity profile. This study evaluated the toxicity profile of DL/D/P by assessing impact on kidney function and structure of healthy and P. berghei-infected mice. The kidney been a major organ for the excretion of drugs increases its vulnerability to toxicity (Craig et al., 2014). The assessments of organ and body weights are essential aspects in the toxicity studies of chemical substances (Sellers et al., 2007). In this study, DL/D/P had no deleterious impacts on the body and kidney weights of parasitized mice. However, DL/D/P increased kidney weight and decreased body weight in healthy mice after 28days of treatment. The decreased body weight and increased kidney weight may be due to appetite suppression and inflammation caused by DL/D/P, respectively. Renal biomarkers are used to envisage the extent of kidney injury. Clinically, renal biomarkers give essential information on renal status during therapeutic interventions using pharmacologic agents (Gowda et al., 2010). In this study, serum electrolytes, creatinine, uric acid and urea were assessed (Adikwu et al., 2019a) to ascertain the impact of DL/D/P on renal function of treated mice. Treatment with DL/D/P had no detrimental effects on serum creatinine, urea and uric acid levels of parasitized mice. In contrast, DL/D/P treatment for 28 days in healthy mice, increased serum creatinine, urea and uric acid levels. The observations in healthy mice are signs of nephrotoxicity (Adikwu et al., 2019b). Electrolytes such as sodium, potassium, bicarbonate and chloride are vital for basic functions including the generation and conduction of action potentials in nerves and muscles (Shrimanker and Bhattarai, 2021). Serum electrolyte test is used to examine acid-base imbalance, which is important in clinical conditions associated with renal, endocrine and other systems (Gowda et al., 2010). In this study, DL/D/P had no detriment impact on serum electrolytes of healthy and parasitized mice. Reactive oxygen species (ROS) such as superoxide radicals, hydrogen peroxide and hydroxyl radicals are produced by mitochondria in both physiological and pathological state as metabolic by-products (Pizzino et al., 2017). When ROS increases beyond regulation, it becomes harmful to important cellular structures (proteins, lipids, and nucleic acids) (Wu et al., 2013). Cells utilize antioxidant defensive mechanism like GSH, SOD, CAT, and GPx, to prevent damage due to ROS-induced oxidative stress. Oxidative stress establishes itself when there is excess production and accumulation of ROS in cells which overwhelms antioxidant capacity (Pizzino et al., 2017). In this present study, there were no alterations in kidney antioxidants of parasitized mice treated with DL/D/P for 4 days. However, decreased kidney antioxidants were noted in healthy mice treated with DL/D/P for 28 days. The observation in healthy mice is a pointer to oxidative stress (Adikwu et al., 2021). Lipid peroxidation is a free-radical-mediated chain of reactions that once initiated, results in an oxidative deterioration of polyunsaturated lipids. MDA is a low-molecular weight aldehyde that is produced during lipid peroxidation (Grotto et al., 2009). The determination of MDA has attracted interest, because it offers a facile means of assessing lipid peroxidation in biological systems. The current study observed normal Kidney MDA levels in parasitized mice treated with DL/D/P for 4 days. But kidney MDA levels were elevated in mice treated with DL/D/P for 28 days. The observation connotes that DL/D/P might have induced kidney lipid peroxidation through the degeneration of poly unsaturated fatty acids due to ROS generation in healthy mice. Histology, an imperative tool in toxicity studies is a descriptive and interpretive science that examines the structural manifestations of disease at the light-microscopic level (Crissman et al., 2004). In this study, the histologic examination of the kidneys of DL/D/P treated healthy mice showed tubular necrosis, enlarged bowman's space and vacuolated mesangial cells in the glomerulus. This may be due to oxidative stress induced by DL/D/P causing damage to cellular components (protein, lipids and DNA) in the kidney. In this study, altered serum renal biochemical markers and kidney histology in D/P treated healthy mice support previous studies (Olayinka and Ore, 2013). Despite the fact that DL seems safe (Monroe et al., 2003), this study observed renal damage marked by elevated serum creatinine, urea, uric acid and vacuolated mesangial cells in the glomerulus of treated healthy mice.

5. CONCLUSION

This study suggests that the use of DL/D/P as an antimalarial drug may not perturb the kidney, but prolonged use may cause nephrotoxicity.

ACKNOWLEDGEMENT

The authors appreciate the staff of Pharmacology Laboratory, Faculty of Basic Clinical Sciences, University of Port Harcourt, Rivers State, Nigeria.

Funding:

This study did not receive any external funding.

Conflict of Interest:

The authors declare that there are no conflicts of interests.

Ethical approval & declaration

In this article, the animal regulations followed as per the ethical committee guidelines of Department of Pharmacology/Toxicology, Faculty of Pharmacy, Niger Delta University, Bayelsa State, Nigeria; the authors observed the renal impact of deslorated dihydroartemisinin /piperaquine on healthy and parasitized mice. The Animal ethical guidelines are followed in the study for observation, identification & experimentation.

Data and materials availability:

All data associated with this study are present in the paper.

REFERENCES

- 1. Adikwu E, Biradee I, Ogungbaike TO. Therapeutic Benefit of resveratrol on 5-fluorouracil-induced nephrotoxicity in rats. *J Biomed Res* 2019; 6 (2) 11-16b
- Adikwu E, Ebinyo NC, Amgbare BT. Protective Activity of Selenium against 5-Fluorouracil-Induced nephrotoxicity in Rats. Cancer Trans Res 2019;5(3):50–5a
- 3. Aebi H. Catalase in vitro. Methods Enzymol 1984; 105:121-6.
- Alabi MA, Kareem FA, Akinwunmi F, Obatoye AO. Nephrotoxicity of Therapeutic Dose of Dihydroartemisinin-Piperaquine Phosphate in Male and Female Animals. *J Prog Res in Modern Phys and Chem.* 2018 (3), 121-127
- Al-nouri ZL, Reese JA, Terrell DR, Vesely SK, George JN, Drug-induced thrombotic microangiopathy: a systematic review of published reports. *Blood* 2015, 125; 616-9
- Aneesa, S. (2011). Evaluation of antihistamines for in vitro antimalarial activity against Plasmodium falciparum. https://api.semanticscholar.org/CorpusID:82705763.
 Accessed 29 October 2020
- Azade Sari (2019). Nephrotoxic Effects of Drugs, Poisoning in the Modern World - New Tricks for an Old Dog?, Ozgur Karcioglu and Banu Arslan, IntechOpen. Available from: https://www.intechopen.com/books/poisoning-in-the-mode rn-world-new-tricks-for-an-old-dog-/nephrotoxic-effects-ofdrugs
- Bhattacharya, U., Roy, S., Kar, P. K., Sarangi, B. & Lahiri, S.
 C. Histamine & kinin system in experimental malaria. *Indian J Med Res* 1998, 88, 558–563.
- 9. Buege JA, Aust SD. Microsomal lipid peroxidation. Meth Enzymol 1978; 52:302-10.
- Campos SB, Rouch LH, Seguro AC. Effects of sodium artesunate, a new antimalarial drug, on renal function. *Kidney Int.* 2001; 59:1044-51.
- 11. Craig, EA, Yan, Z, and Zhao, QJ. The relationship between chemical-induced kidney weight increases and kidney histopathology in rats. *J. Appl. Toxicol.*, 2015. 35, 729–736.
- 12. 12. Crissman JW, Goodman DG, Hildebrandt PK, Maronpot RR, Prater DA, Riley JH et al Best Practices

- Guideline: Toxicologic Histopathology. *Toxicol Path*, 32:126–131, 2004
- 13. Dhodi DK, Bhagat SB, Pathak D, Patel SB. Drug-induced nephrotoxicity. *Intern J Basic & Clin Pharm*. 2014; 3(4):591-597.
- 14. Gbotosho, G.O., Happi, C.T., Ganiyu, A. *et al.* Potential contribution of prescription practices to the emergence and spread of chloroquine resistance in south-west Nigeria: caution in the use of artemisinin combination therapy. *Malar J* 2009; 8, 313
- 15. Georgewill UO, Ebong NO, Adikwu E. Antiplasmodial activity of desloratadine-dihydroartemisinin-piperaquine on Plasmodium berghei infected mice. *J Appl Biol* and Biotech 2021; 9(2):169-173.
- 16. Gounden V, Bhatt H, Jialal I. Renal Function Tests. In: StatPearls [Internet]. Treasure Island (FL). https://www.ncbi.nlm.nih.gov/books/NBK507821/
- 17. Gowda, S., Desai, P. B., Kulkarni, S. S., Hull, V. V., Math, A. A., & Vernekar, S. N. Markers of renal function tests. *North Amer J Med Sci*, 2010. 2(4), 170–173.
- 18. Kazmi F, Barbara JE, Yerino P, Parkinson A. A long-standing mystery solved: the formation of 3-hydroxydesloratadine is catalyzed by CYP2C8 but prior glucuronidation of desloratadine by UDP-glucuronosyltransferase 2B10 is an obligatory requirement. *Drug Metab Dispos.* 2015 43(4):523-33.
- 19. Li Q, Xie LH, Johnson TO, Si Y, Haeberle AS, Weina PJ. Toxicity evaluation of artesunate and artelinate in Plasmodium berghei-infected and uninfected rats. Trans R Soc *Trop Med Hyg.* 2007; 101:104-12.
- 20. Liles NW, Page EE, Liles, AL, Vesely SK, Raskob GE, George JN Diversity and severity of adverse reactions to quinine: a systematic review *Am J Hematol*, 2016, 91; 461-6
- 21. Lillie M A Barnett, Brian S Cummings, Nephrotoxicity and Renal Pathophysiology: A Contemporary Perspective, *Toxicol Sci*, 2018; 164 (2) 379–390

- 22. Maegraith, B. Fletcher, A. The pathogenesis of mammalian malaria. *Adv Parasitol* 10, 49–75 (1972)
- 23. Mahmoudi J, Sadigh-Eteghad S, Salehi-Pourmehr H, Gharekhani A, Ziaee M. Nephrotoxicity of Chloroquine and Hydroxychloroquine in COVID-19 Patients *Adv Pharm Bull*, 2021, 11(1), 1-2
- 24. Monroe, E., Finn, A., Patel, P., Guerrero, R., Ratner, P., Bernstein, D., & Desloratadine Uritcaria Study Group. Efficacy and safety of desloratadine 5 mg once daily in the treatment of chronic idiopathic urticaria: a double-blind, randomized, placebo-controlled trial. *J Amer Acad of Derm*, 2003; 48(4), 535–541.
- 25. Musabayane, C.T, Copper, R.G, Prasada Rao, P.V.V, Balen, R.J. Effects of ethanol on the changes in renal fluid and electrolyte handling and kidney morphology induced by long term chloroquine induced administration to rats. *Alcohol* 2000, 22, 129–138.
- 26. Oduola AMJ, Sowunmi A, Milhous WK, Brewer TG, Kyle DE, Gerena L, Rossan RN, Salako LA, Schuster B: In vitro and in vitro reversal of chloroquine resistance in Plasmodium falciparum with promethazine. Am J Trop Med Hyg. 1998, 58: 625-629.
- 27. Olayinka ET, Ore A. Alterations in Antioxidant Status and Biochemical Indices Following Administration of Dihydroartemisinin-Piperaquine Phosphate (P-ALAXIN®). *J Pharm and Biol Sci* 2013 (5)4 43-53
- 28. Omitowoju GO, Ogundahunsi OAT, Milhous WK, Gerena L, Sowunmi A, Schuster BG, Oduola AMJ: Chlorpheniramine: a resistance reversing agent with potential clinical application. *Am J Trop Med Hyg.* 1992, 47, 175
- 29. Pazhayattil, G. S., Shirali, A. C. Drug-induced impairment of renal function. *Intern J Nephrol and Renovasc dis*, 2014; 7, 457–468.
- Peters W, Ekong R, Robinson BL, Warhurst DC, Pan XQ: Antihistaminic drugs that reverse chloroquine resistance in *Plasmodium falciparum*. *Lancet ii*. 1989, 334-335.
- Pizzino G, Irrera N, Cucinotta M, Pallio G, Mannino F, Arcoraci V, Squadrito F, Altavilla D, Bitto A. Oxidative Stress: Harms and Benefits for Human Health. Oxid Med Cell Longev. 2017; 8416763.
- 32. Reuter SE, Evans AM, Shakib S, Lungershausen Y, Francis B, Valentini G, Bacchieri A, Ubben D, Pace S. Effect of food on the pharmacokinetics of piperaquine and dihydroartemisinin. *Clin Drug Invest*. 2015; 35(9):559-67.
- 33. Rotruck JT, Pope AL, Ganther HE, Swanson AB, Hafeman DG, Hoekstra WG. Selenium: biochemical role as a component of glutathione peroxidase. *Sci* 1973; 179:588-90.
- 34. Sedlak J, Lindsay RH. Estimation of total, protein-bound, and nonprotein sulfhydryl groups in tissue with Ellman's reagent. Anal Biochem 1968; 25:192-205.

- 35. Sellers RS, Mortan D, Michael B, et al. Society of Toxicologic Pathology Position Paper: Organ Weight Recommendations for Toxicology Studies. *Toxicol Path.* 2007; 35(5):751-755.
- Shrimanker I, Bhattarai S. Electrolytes. [Updated 2021 Jul 26]. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2021 Jan. Available from: https://www.ncbi.nlm.nih.gov/books/NBK541123/
- 37. Srichaikul, T., Archararit, N., Siriasawakul, T. & Viriyapanich, T. Histamine changes in Plasmodium falciparum malaria. *Trans R Soc Trop Med Hyg* 1976; 70, 36-38.
- 38. Sun M, Zigma S. An Improved spectrophotometer assay of superoxide dismutase based on epinephrine antioxidation. *Anal Biochem* 1978; 90:81-9.
- 39. Sun XD, Zhang ZX, Wang J, Deng Y, Yang YC, Lasi JH, Sun XY, Wang H. Therapeutic efficacy and safety of compound dihydroartemisinin/piperaquine for uncomplicated Plasmodium falciparum infection in Laiza City of Myanmar bordering on China. *Zhongguo Ji Sheng Chong Xue Yu Ji Sheng Chong Bing Za Zhi*. 2011; 29(5):372-5.
- 40. Wiwanitkit V. Antimalarial drug and renal toxicity. *J Nephropharmacol.* 2015; 5(1):11-12. Published 2015 Dec 23.
- 41. World Health Organization. Guidelines for the treatment of malaria. 2015. http://apps.who.int/iris/bitstream/10665/16244 1/1/9789241549127_eng.pdf?ua=1. Accessed 27 July 2021.
- 42. Wu JQ, Kosten TR, Zhang XY, Free radicals, antioxidant defense system, and schizophrenia, *Progress in Neuro-Psychopharm & Biol Psych*, 2013; 46, 200–206.